0957-4166/97 \$17.00 + 0.00



PII: S0957-4166(97)00344-3

Total synthesis of both enantiomers of trans-β-hydroxypipecolic acid

Lucia Battistini, Franca Zanardi*, Gloria Rassu, Pietro Spanu, Giorgio Pelosi, Giovanna Gasparri Fava, Marisa Belicchi Ferrari and Giovanni Casiraghi* A.*

^a Dipartimento Farmaceutico dell'Università, Viale delle Scienze, I-43100 Parma, Italy
 ^b Istituto per l'Applicazione delle Tecniche Chimiche Avanzate del CNR, Via Vienna 2, I-07100 Sassari, Italy
 ^c Dipartimento di Chimica Generale ed Inorganica, Chimica Analitica, Chimica Fisica dell'Università, Viale delle Scienze, I-43100 Parma, Italy

Abstract: trans-β-Hydroxypipecolic acids of both L- and D-series, L-1 and D-1, have been straightforwardly prepared in 14% and 15% yields, respectively, starting from glyceraldehyde imines D-7 and L-7 as useful three-carbon chirons. The key feature of these parallel syntheses lies on the highly diastereoselective character of the initial coupling manoeuver between silyloxy furan TBSOF and imines 7, which ultimately accounts for the relative, and hence absolute configuration of the target pipecolic acids. © 1997 Elsevier Science Ltd

Introduction

3-Hydroxypipecolic acids, six-membered cyclic α -amino- β -hydroxy acids [e.g. (2S,3S)-isomer L-1, Figure 1], constitute non-natural variants of a structural motif often encountered in a variety of functional molecules—e.g. naturally occurring (2S,4R)-4-hydroxypipecolic acid 2,¹ the uronic acid derivative of deoxynojirimycin 3,² and potent β -glucuronidase inhibitor D-glucaro- δ -lactam 4³—and may be regarded as an "expanded" hydroxylated proline (e.g. 5), or a conformationally restricted serine derivative (e.g. 6).

As a consequence, a practical route providing an easy access to these multifunctional, chiral substances in a structurally defined form should be highly desirable,⁴ also considering that these compounds might be precious scaffolds to be incorporated into conformationally restricted peptidomimetics of biological relevance.

Results and discussion

Planning

A variable plan addressing the synthesis of a given 3-hydroxypipecolic acid of type A is illustrated in Scheme 1. Here, the piperidine ring of the target amino acid A, derived from

Figure 1.

^{*} Corresponding author. Email: casirag@ipruniv.cce.unipr.it

piperidinone **B** by oxidative fission of the C-1'-C-2' linkage and C-6 carbonyl reduction, is built up via γ -lactone-to- δ -lactam expansion ($C \rightarrow B$). Lactone **C** is obtained by saturation and selective deprotection of butenolide **D**, which is, in turn, available by Lewis acid-catalyzed coupling of 2-[(tert-butyldimethylsilyl)oxy]furan (hereafter TBSOF) with properly protected glyceraldehyde imine **E**. Accordingly, while TBSOF represents the nucleophilic synthon supplying for the hydroxylated C-3-C-6 portion of the pipecolic acid structure (bold lines in Scheme 1), glyceraldehyde imine **E** may be viewed as a (chiral) glycine cation surrogate.⁵

PG = suitable protective groups

Scheme 1.

The asymmetric induction issue—i.e. the selective formation of a given structurally defined isomer—can be addressed by controlling the stereochemical course of the crucial coupling step (TBSOF+ $E\rightarrow D$), wherein two stereocenters (C-4 and C-5) are newly generated. Thus, the *syn* versus *anti* configuration of the C-4 and C-5 carbon atoms within **D** will dictate the relative stereodisposition of the carboxyl and hydroxyl components embodied in **A** (*cis* versus *trans*), while the single chiral element within **E** will control the absolute configuration of the target (facial diastereoselection).

Synthesis of trans-3-hydroxypipecolic acids L-1 and D-1

The stereoselective synthesis of pipecolic acid L-1 commenced with the key coupling reaction between 2-silyloxyfuran TBSOF and 2,3-O-isopropylidene-D-glyceraldehyde N-benzyl imine (D-7) (Scheme 2).⁶ Treatment of freshly prepared imine D-7 with TBSOF in the presence of 0.5-0.6 molar equivalents of TBSOTf⁷ in CH₂Cl₂ at -80°C gave rise to 4,5-anti-5,6-anti-configured butenolide 8 as the predominant isomer, along with a small amount of the 4,5-anti-5,6-syn-disposed diastereoisomer 98 (90:10 dr, 90% combined yield). The stereochemical assignment of the major lactone 8 was only tentative at this point, based on chiroptical considerations, but its structure is firmly ascertained at a later stage of the synthesis (vide infra). Conveniently, the 9:1 crude mixture of butenolides 8 and 9 was subjected to catalytic hydrogenation (Pd/C, THF, NaOAc), which provided saturation of the double bond with concomitant removal of the benzyl protective group. There was obtained crystalline amino lactone 10 in 75% yield, after easy flash chromatographic purification.

At this juncture, furanone-to-piperidinone expansion had to be performed, and this was readily achieved by treating compound 10 with neat 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) at 80°C. Pure piperidinone 11 was obtained in a good 82% yield, which was cleanly transformed to crystalline piperidine 12 (70% yield) by alane reduction at -80°C (LiAlH₄, AlCl₃, THF). ¹⁰

Scheme 2.

The assignment of the relative *trans*-disposition of the substituents at the C-4 and C-5 positions within 12 followed from ¹H NMR spectral analysis that showed a diagnostic coupling constant between the H-4 and H-5 protons ($J_{4,5}$ =8.8 Hz), thus confirming a *trans*-diaxial configuration for these nuclei. Unambiguous relative and absolute configuration of 12 was confirmed by a single crystal X-ray analysis (*vide infra*), thus corroborating the structures of all the related products in Scheme 2.

Having the hydroxylated piperidine moiety at hand, all that remained was the unmasking of the side diol appendage to a carboxylic function. Thus, protection of both NH and OH groups in 12 was first carried out by acetylation (Ac₂O, pyridine, DMAP), furnishing N,O-bis-acetylated compound 13 in 90% yield. Notably, the ¹H and ¹³C NMR spectra in CDCl₃ at room temperature showed compound 13 to be a 1:1 mixture of two isomeric components, probably arising from amide isomerism.

Fully protected piperidine 13 was readily transformed into aldehyde 14 by a two-step procedure involving acidic deprotection (80% aq. AcOH, 60°C), followed by aqueous sodium periodate oxidation (68% yield for the two steps to 14). Crude aldehyde 14 was directly oxidized to protected pipecolic acid 15 (RuO₂·2H₂O, solid NaIO₄, 78% yield), which was finally converted to pipecolic acid L-1 by a two-step deprotective protocol involving NaOMe treatment in methanol to furnish a mono N-acetyl intermediate, followed by exposure to 6 N aq. HCl at 80°C and chromatographic purification. (2S,3S)-3-Hydroxypipecolic acid (L-1) was recovered as an off-white solid material (mp 230–238°C, decomp.) in 77% yield from 15, which corresponds to 14% overall yield for the entire ten-step sequence from D-7.

Very conveniently, (2R,3R)-3-hydroxypipecolic acid D-1 was prepared by starting from 2,3-0-isopropylidene-L-glyceraldehyde N-benzyl imine L-7 and TBSOF through intermediates ent-8-ent-15 by just paralleling the synthetic procedure described above for its enantiomer L-1 (Scheme 3). There was obtained pipecolic acid D-1 (D-series) as a white solid material (mp 234-239°C, decomp.) in 15% overall yield for the whole sequence from L-7.

Scheme 3. (See Scheme 2 for reaction conditions.)

Mechanistic considerations

The diastereoselective behavior of the opening move in this synthesis, i.e. the vinylogous Mukaiyama-aldol addition¹¹ of TBSOF to chiral imine derivatives, solely dictates the generation of all the stereocenters within the target compounds and the related intermediates. Irrespective of the nature of the Lewis acid employed, the reaction strongly favored the formation of the 4,5-anti-5,6-anti-configured adducts (8 and ent-8), being the optimal conditions observed when a catalytic amount of TBSOTf (0.5-0.6 equivalents) was used.⁷

Of note, this behavior markedly differs from the general outcome of the Lewis acid-assisted reactions involving TBSOF (and related silyloxy furan, pyrrole, and thiophene reagents) and aldehyde electrophiles, which lead to 4.5-syn-5.6-anti-configured adducts, preferentially.⁶ Although a complete rationale accounting for the observed stereochemical behavior is hard to construct, one can formulate a mechanistic path as that outlined in Scheme 4, accounting for the preferential formation of the 4.5-anti-adducts II (for α -alkoxyimine derivatives) and 4.5-syn-adducts III (for α -alkoxyaldehydes).

For aldehydes, a Diels-Alder like transition state TS3 leading to compounds III seems to be preferred over TS4 (open chain), whilst for imine derivatives, an open chain model of type TS2, giving rise to adducts II, appears more favorable over TS1. Likely, TS2 seems to be stabilized over TS1 due to diminished steric congestion, whereas TS3 is advantaged over TS4 owing to favorable orbital overlapping and less severe congestion. Instead, the facial diastereoselection governing the 5,6-relative configuration of the adducts is substrate-independent, with 5,6-anti-configured adducts (Felkin type approach) favored using both aldehyde and imine substrates.

X-Ray structural analysis

An ORTEP plot of compound 12 with its numbering scheme is displayed in Figure 2. The absolute configuration of the stereocenters has been assigned as 2S, 1S, 6S (crystallographic numbering; 4S, 5S, 6S elsewhere) based on the chirality of the related precursor D-7. The overall conformation of the molecule is determined by one intramolecular interaction^{12a,b} between C6-H6 and O1 [2.910(8) Å] and by an intramolecular hydrogen bond N1-H1N···O2 [2.835(7) Å].

The orientation of the O1-H group with respect to the dioxolane ring is shown by the following torsion angle C6C1C2O1=54.4(7)°. The H1···H2, H6···H1A and H1A···H1N distances are 2.61, 2.31, and 2.23 Å, respectively, with torsion angles of -175.7, 60.7, and 71.4° in antiperiplanar, synclinal, and synclinal conformation. As expected, the arrangement of H2 and H1A atoms is *trans*-

diaxial with a torsion angle H2-C2-C1-H1A of 178.5°, in agreement with the ¹H NMR measurements (vide supra).

Scheme 4.

The piperidine ring is in the $^{1}C_{4}$ chair conformation [puckering parameters: q_{2} =0.056(7) Å, q_{3} =-0.563(7) Å, φ_{2} =174.1(7.0)°, Q=0.565(7) Å, Θ_{2} =174.3(7)°], 12c as unprotected nojirimycin 12d and other simple piperidine derivatives. 12e The dioxolane ring shows a conformation closest to twist with a twofold axis passing through the C6 atom [puckering parameters: q_{2} =0.294(6) Å, φ_{2} =78.3(1.2)°]. 12c,f The dihedral angle between the dioxolane ring and the mean plane of the piperidine is 96.5(2)°. Bond lengths and angles in the piperidine and dioxolane rings are in good agreement with the values reported for similar moieties. 12e,g The packing is mainly governed by O1-H1 · · · N1(x,y,z+1) hydrogen bond of 3.033(8) Å with the O-H· · · N angle of 165.3(4)°. A weak interaction C7-H7B · · · O1(x,y,z-1)=3.47(1) Å is also present, so chains of molecules run along the crystallographic c axis (Figure 3).

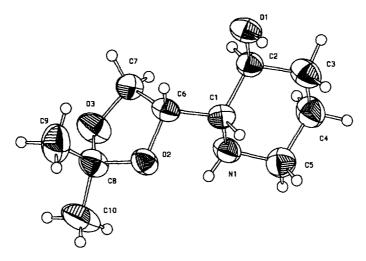


Figure 2. ORTEP view of the molecule 12 in its correct absolute configuration showing the atomic numbering scheme.

Thermal ellipsoids enclose 50% of probability and the hydrogen atoms are drawn with an arbitrary diameter.

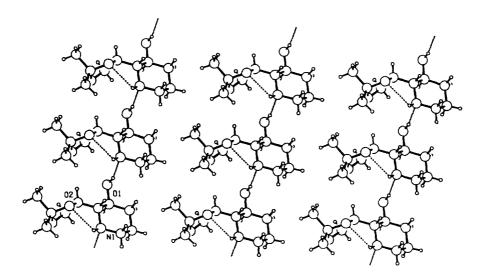


Figure 3. Projection of the structure on the (010) plane showing the most significant hydrogen bonds.

Conclusion

Practical methodology for the synthesis of enantiopure (2S,3S)- and (2R,3R)-3-hydroxypipecolic acids L-1 and D-1 has been presented, that exploits glyceraldehyde imines D-7 and L-7 as synthetic equivalents of chiral α -glycine cation. The highly diastereoselective and regioselective character of the key operation, namely the vinylogous aldol addition of silyloxy furan TBSOF to imines 7, dictated the formation of *trans*-disposed 3-hydroxy pipecolic acids, while imparting chirality into the targets and related intermediates. The high yields of each reaction step and the availability of almost all the isolated intermediates as crystalline solids render this protocol an efficient way to access these rare amino acids on both small and large scales.

The potentiality of the chemistry here disclosed overrides the production of simple pipecolic acids of type 1, being certain synthetic intermediates envisioned as valuable building blocks en route to a variety of multifunctional structures of biological relevance. As an example, proper dihydroxylation

of the double bond within 8 could secure preparation of piperidinose sugars of the nojirimycin family, while enolate-driven α -alkylation of 11 would pave the way to acyclic hydroxyethylene dipeptide isosteres, a promising progeny of aspartic protease inhibitors.

Experimental

General procedures

Melting points were determined (uncorrected) on an Electrothermal apparatus. Optical rotations were measured on a Perkin-Elmer 241 polarimeter and are given in units of 10⁻¹ deg cm² g⁻¹. The ¹H (300 MHz) and ¹³C (75 MHz) NMR spectra were recorded using a Bruker AC-300 spectrometer. Chemical shifts are quoted in parts per million (δ) relative to tetramethylsilane (0.0 ppm) or to the specific solvent as an internal standard, and coupling constants (*J*) are measured in hertz (Hz). Analytical thin-layer chromatography was carried out on Merck Kieselgel 60 F₂₅₄ glass-backed plates. Flash chromatography was performed on 32–63 μm silica gel (ICN Biomedicals), using the reported solvent mixtures. The compounds were visualized by dipping the plates in an aqueous H₂SO₄ solution of cerium sulfate and ammonium molybdate, followed by heating. All the solvents were dried according to common methods and distilled before use. All the reactions were carried out under an oxygenfree nitrogen atmosphere. Elemental analyses were performed by the Microanalytical Laboratory of University of Sassari.

Materials

2-[(tert-Butyldimethylsilyl)oxy]furan (TBSOF) was prepared on a multigram scale from 2-furaldehyde (Aldrich) according to a described protocol.^{6d} 2,3-O-Isopropylidene-D- and L-glyceraldehyde N-benzylimines (D-7 and L-7) were obtained by reacting benzylamine (Aldrich) and 2,3-O-isopropylidene-D- and L-glyceraldehyde, respectively, in CH₂Cl₂, in the presence of anhydrous MgSO₄. The crude materials were used as such in the subsequent coupling processes.

5-(N-Benzylamino)-6,7-O-isopropylidene-2,3,5-trideoxy-D-ribo-hept-2-enonic acid 1,4-lactone 8 and 5-(N-benzylamino)-6,7-O-isopropylidene-2,3,5-trideoxy-D-lyxo-hept-2-enonic acid 1,4-lactone 9

To a stirring solution of imine D-7 (4.55 g, 20.64 mmol) in anhydrous CH_2Cl_2 (80 mL) under nitrogen atmosphere cooled to $-80^{\circ}C$ were sequentially added TBSOF (4.09 g, 20.6 mmol) and TBSOTf (2.37 mL, 10.32 mmol), and the resulting mixture was allowed to react for 3 h at $-80^{\circ}C$. The reaction was then quenched at the same temperature by the addition of saturated aqueous NaHCO₃ and, after ambient temperature was reached, the mixture was extracted with CH_2Cl_2 (3×20 mL). The combined organic layers were washed with brine (2×15 mL), dried with MgSO₄, filtered, and evaporated to give an oily yellowish crude residue (5.64 g, 90%), whose ¹H NMR spectrum revealed the presence of a ~9:1 mixture of isomeric 8 and 9. While almost all the recovered residue was used as such in the subsequent hydrogenation step, a small portion (0.60 g) was subjected to flash chomatographic purification (hexanes/EtOAc 60:40), in order for the predominant lactone 8 to be fully characterized. There was obtained pure 8 (498 mg, 83%), along with 54 mg (9%) of the isomeric product 9.

Compound **8**: white crystals, mp 66–67°C; R_f =0.37 (hexanes/EtOAc 60:40); $[\alpha]_D^{20}$ =-35.2 (c=0.8, CHCl₃); 1 H NMR (300 MHz, CDCl₃): δ 7.53 (dd, J=5.8, 1.6 Hz, 1H), 7.28 (m, 5H), 6.16 (dd, J=5.8, 2.1 Hz, 1H), 5.35 (app. quint., J=2.1 Hz, 1H), 4.07 (dd, J=8.3, 6.3 Hz, 1H), 3.95 (m, 1H), 3.7–3.9 (m, 4H), 3.08 (dd, J=7.5, 4.2 Hz, 1H), 1.39 (s, 3H), 1.31 (s, 3H); 13 C NMR (75 MHz, CDCl₃) δ 172.7, 154.3, 139.5, 128.1 (5C), 122.2, 109.6, 83.6, 75.5, 67.5, 61.2, 52.9, 26.5, 25.0. Anal. Calcd for $C_{17}H_{21}NO_4$; C, 67.31; H, 6.98; N, 4.62. Found: C, 67.04; H, 7.15; N, 4.50.

5-(N-Benzylamino)-6,7-O-isopropylidene-2,3,5-trideoxy-L-ribo-hept-2-enonic acid 1,4-lactone ent-8 and 5-(N-benzylamino)-6,7-O-isopropylidene-2,3,5-trideoxy-L-lyxo-hept-2-enonic acid 1,4-lactone ent-9

The title compounds were prepared according to the synthetic procedure described for their enantiomers 8 and 9, by employing 2,3-O-isopropylidene-L-glyceraldeyde N-benzylimine L-7 in place of D-7. Starting with TBSOF (3.57 g, 18.0 mmol), there were obtained 4.8 g (87%) of a crude oily residue containing a 9:1 mixture of ent-8 and ent-9, which was used as such in the next hydrogenation step. A small amount of this material (0.6 g) was flash chromatographed (hexanes/EtOAc 60:40), furnishing 450 mg (75%) of pure ent-8, along with 50 mg (8%) of pure ent-9.

Compound *ent-8*: white crystals, mp 64–65°C; R_f =0.37 (hexanes/EtOAc 60:40); $[\alpha]_D^{20}$ =+36.0 (c=0.8, CHCl₃); The ¹H NMR and ¹³C NMR spectra coincided with those of its enantiomer 8. Anal. Calcd for $C_{17}H_{21}NO_4$: C, 67.31; H, 6.98; N, 4.62. Found: C, 67.34; H, 7.07; N, 4.56.

5-Amino-6,7-O-isopropylidene-2,3,5-trideoxy-D-ribo-heptonic acid 1,4-lactone 10

To a stirring solution of crude material 8+9 (5.0 g, 16.48 mmol) in anhydrous THF (150 mL) was added 10% Pd on carbon (0.5 g) and a small amount of sodium acetate (120 mg) at room temperature. The reaction vessel was evacuated and thoroughly purged with hydrogen (three times), and the resulting heterogeneous mixture was stirred under a balloon of hydrogen for 24 h. After hydrogen evacuation, the catalyst was filtered off, and the filtrate was concentrated under vacuum to afford a crude oily residue that was subjected to flash chromatographic purification (EtOAc/MeOH 90:10). Pure amino lactone 10 (2.7 g) was recovered in 75% yield as a white crystalline solid: mp 62–65°C, R_f =0.3 (EtOAc/MeOH 90:10); $[\alpha]_D^{20}$ =+19.1 (c=0.6, CHCl₃); 1 H NMR (300 MHz, CDCl₃) δ 4.59 (ddd, J=8.0, 7.3, 4.7 Hz, 1H), 4.07 (dd, J=7.7, 6.2 Hz, 1H), 3.99 (ddd, J=6.9, 6.2, 5.9 Hz, 1H), 3.88 (dd, J=7.7, 5.9 Hz, 1H), 3.24 (dd, J=6.9, 4.7 Hz, 1H), 2.54 (m, 2H), 2.21 (m, 2H), 2.01 (bs, 2H), 1.40 (s, 3H), 1.32 (s, 3H); 13 C NMR (75 MHz, CDCl₃) δ 176.9, 109.2, 81.0, 75.6, 66.4, 54.8, 28.5, 26.4, 25.0, 22.1. Anal Calcd for $C_{10}H_{17}NO_4$: C, 55.80; H, 7.96; N, 6.51. Found: C, 55.92; H, 8.07; N, 6.40.

5-Amino-6,7-O-isopropylidene-2,3,5-trideoxy-L-ribo-heptonic acid 1,4-lactone ent-10

Saturated amino lactone *ent-***10** was prepared from crude mixture *ent-***8**+*ent-***9** (4.2 g, 13.84 mmol), by adopting the above hydrogenation procedure to **10**. After flash chromatographic purification (EtOAc/MeOH 90:10) there were obtained 2.3 g (78%) of pure *ent-***10** as white crystals: mp 62–63°C; R_f =0.28 (EtOAc/MeOH 90:10); $[\alpha]_D^{20}$ =-18.5 (c=0.5, CHCl₃). The ¹H and ¹³C NMR spectra coincided with those reported for its enantiomer **10**. Anal. Calcd for $C_{10}H_{17}NO_4$: C, 55.80; H, 7.96; N, 6.51. Found: C, 55.89; H, 7.79; N, 6.35.

5-Amino-6,7-O-isopropylidene-2,3,5-trideoxy-D-ribo-heptonic acid 1,5-lactam 11

2.7 g (12.5 mmol) of amino lactone **10** were dissolved in 10 mL of DBU and the resulting mixture was warmed to 80°C for 4 h. The solution was then concentrated under vacuum leaving a brown residue that was purified by flash chromatography eluting with EtOAc/MeOH (75:25). Pure lactam **11** was obtained (2.2 g, 82%) as a white crystalline solid: mp 110–113°C; R_f=0.6 (EtOAc/MeOH 75:25); $[\alpha]_D^{20}$ =+22.3 (c=0.8, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 6.47 (bs, 1H), 4.50 (bs, 1H), 4.23 (app. q, J=6.4, 6.3, 6.1 Hz, 1H), 4.06 (dd, J=8.4, 6.4 Hz, 1H), 3.79 (dd, J=8.4, 6.3 Hz, 1H), 3.75 (m, 1H), 3.42 (td, J=6.5, 1.4 Hz, 1H), 2.46 (dt, J=18.0, 5.8 Hz, 1H), 2.29 (ddd, J=18.0, 9.5, 6.3 Hz, 1H), 1.98 (m, 1H), 1.86 (m, 1H), 1.41 (s, 3H), 1.32 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.2, 109.5, 76.3, 65.8, 65.2, 59.4, 28.1, 27.4, 26.2, 24.8. Anal. Calcd for C₁₀H₁₇NO₄: C, 55.80; H, 7.96; N, 6.51. Found: C, 55.74; H, 8.02; N, 6.60.

5-Amino-6,7-O-isopropylidene-2,3,5-trideoxy-L-ribo-heptonic acid 1,5-lactam ent-11

The title compound was prepared employing 2.3 g (10.68 mmol) of γ -lactone ent-10 by following the procedure described for its enantiomer 11. There were obtained 1.84 g (80%) of pure lactam ent11 as a white crystalline solid: mp 108-110°C; R_f =0.6 (EtOAc/MeOH 75:25); $[\alpha]_D^{20}$ =-20.8 (c=0.8,

CHCl₃). The 1 H and 13 C NMR spectra coincided with those of its enantiomer 11. Anal. Calcd for $C_{10}H_{17}NO_4$: C, 55.80; H, 7.96; N, 6.51. Found: C, 55.64; H, 8.09; N, 6.38.

6,7-O-Isopropylidene-1,2,3,5-tetradeoxy-1,5-imino-D-ribo-heptitol 12

To a reaction vessel containing anhydrous AlCl₃ (2.0 g, 15.33 mmol) cooled to 0°C under a nitrogen atmosphere, were added 60 mL of dry THF and the resulting colorless solution was stirred at the same temperature for 5 min. Lithium aluminium hydride (46.0 mL of a 1.0 M solution in THF, 46.0 mmol) was then added, and a vigorous bubbling was observed. The colorless AlH₃ solution so obtained was warmed to room temperature and allowed to stir for 20 min, after which time was recooled to -80°C. A pre-cooled (-80°C) solution of lactam 11 (2.2 g, 10.22 mmol) in dry THF (60 mL) was added via syringe to the above alane solution and the resulting cloudy mixture was stirred at -80°C for 1 h, warmed to 20°C, and stirred an additional 20 min. The colorless solution so obtained, once recooled to 0°C, was quenched with MeOH and the resulting slurry was thoroughly extracted with EtOAc (5×30 mL). The combined organic layers were dried (MgSO₄), filtered, and concentrated in vacuum to furnish a colorless crystalline residue that was purified by flash chromatography (EtOAc/MeOH 70:30) providing 1.44 g (70%) of piperidine 12 as a white crystalline solid: mp 112–115°C; R_f =0.32 (EtOAc/MeOH 70:30); $[\alpha]_D^{20}$ =+28.1 (c=0.2, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 4.12 (app. q, J=6.3 Hz, 1H), 4.06 (dd, J=7.7, 6.3 Hz, 1H), 3.90 (dd, J=7.7, 6.3 Hz, 1H), 3.36 (ddd, J=10.3, 8.8, 4.6 Hz, 1H), 2.93 (dquint, J=12.3, 2.0 Hz, 1H), 2.47 (td, J=12.3, 3.1 Hz, 1H), 2.45 (dd, J=8.8, 6.6 Hz. 1H), 2.33 (bs, 2H), 2.03 (m, 1H), 1.65 (m, 1H), 1.40 (s, 3H), 1.33 (s, 3H), 1.1–1.3 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 111.9, 78.8, 67.4, 63.6, 55.2, 45.5, 33.0, 29.6, 26.3, 25.2. Anal Calcd for C₁₀H₁₉NO₃: C, 59.68; H, 9.52; N, 6.96. Found: C, 59.70; H, 9.46; N, 6.90.

6,7-O-Isopropylidene-1,2,3,5-tetradeoxy-1,5-imino-L-ribo-heptitol ent-12

The title compound was prepared according to the above procedure to 12, by employing 1.84 g (8.55 mmol) of *ent-*11. Piperidine *ent-*12 was recovered (1.28 g, 74%) as a white crystalline solid: mp 114–116°C; R_f =0.32 (EtOAc/MeOH 70:30); $[\alpha]_D^{20}$ =-27.0 (c=0.3, CHCl₃). The ¹H and ¹³C NMR spectra were coincident with those of its enantiomeric counterpart 12. Anal. Calcd for $C_{10}H_{19}NO_3$: C, 59.68; H, 9.52; N, 6.96. Found: C, 59.82; H, 9.67; N, 6.79.

4,5-(O,N-Diacetyl)-6,7-O-isopropylidene-1,2,3,5-tetradeoxy-1,5-imino-D-ribo-heptitol 13

To a solution of piperidine 12 (1.0 g, 4.96 mmol) in dry CH₂Cl₂ (50 mL) under nitrogen atmosphere were sequentially added pyridine (2.4 mL, 29.8 mmol), acetic anhydride (1.87 mL, 19.87 mmol), and a catalytic amount of DMAP (50 mg). The mixture was stirred at room temperature for 2 h, quenched with aqueous saturated NaHCO₃ solution, and extracted with CH₂Cl₂ (3×15 mL). The combined organic layers were dried over MgSO₄, filtered, and concentrated in vacuum to provide a crude oily residue that was purified by flash chromatography (EtOAc/MeOH 95:5). There were obtained 1.28 g (90%) of fully protected piperidine 13 as white crystals: mp 85-87°C, R_f=0.4 (EtOAc/MeOH 95:5); $[\alpha]_D^{20} = -50.1$ (c=0.8, CHCl₃); H NMR (300 MHz, CDCl₃, 50:50 mixture of rotamers) δ 5.20 (m, 1H), 4.77 (bd, J=7.0 Hz, 0.5H), 4.61 (bd, J=9.7 Hz, 0.5H), 4.40 (ddd, J=9.2, 5.9, 4.3 Hz, 0.5H), 4.21 (app. q, J=6.5 Hz, 0.5H), 3.95 (dd, J=8.8, 5.9 Hz, 0.5H), 3.94 (dd, J=8.3, 6.5 Hz, 0.5H), 3.82 (dd, J=8.5, 6.5 Hz, 0.5 H), 3.78 (bd, J=10.0 Hz, 0.5H), 3.72 (dd, J=9.0, 4.2 Hz, 0.5H), 3.64 (bd, J=14.0Hz, 0.5H), 3.23 (td, J=13.4, 2.8 Hz, 0.5H), 2.52 (td, J=13.1, 3.1 Hz, 0.5H), 2.11 (s, 3×0.5H), 2.05 (s, 3×0.5 H), 2.04 (s, 3×0.5 H), 1.99 (s, 3×0.5 H), 1.6–1.9 (m, 4H), 1.48 (s, 3×0.5 H), 1.44 (s, 3×0.5 H), 1.33 (s, 3×0.5H), 1.31 (s, 3×0.5H); 13 C NMR (75 MHz, CDCl₃, 50:50 mixture of isomers) δ 171.2, 170.2, 170.0, 168.0, 110.3, 110.0, 74.5, 73.0, 68.0, 67.3, 67.1, 66.4, 59.1, 53.4, 42.9, 37.1, 29.5 (2C), 27.0, 26.3, 25.2 (2C), 24.7, 23.9, 21.8, 21.4, 20.2, 19.2. Anal. Calcd for C₁₄H₂₃NO₅: C, 58.93; H, 8.12; N, 4.91. Found: C, 59.01; H, 8.20; N, 4.83.

4,5-(O,N-Diacetyl)-6,7-O-isopropylidene-1,2,3,5-tetradeoxy-1,5-imino-L-ribo-heptitol ent-13

The above procedure to 13 was followed, by starting with 1.0 g (4.96 mmol) of piperidine *ent*-12. There was obtained pure *ent*-13 (1.3 g, 90%) as a white crystalline solid: mp 86–88°C, R_f =0.4 (EtOAc/MeOH 95:5); $[\alpha]_D^{20}$ =+52.0 (c=0.8, CHCl₃). The ¹H and ¹³C NMR spectra were coincident with those of its enantiomeric counterpart 13. Anal. Calcd for $C_{14}H_{23}NO_5$: C, 58.93; H, 8.12; N, 4.91. Found: C, 59.10; H, 8.28; N, 4.80.

(2S,3S)-1,3-N,O-Diacetyl-2-formyl-3-hydroxypiperidine 14

Protected piperidine 13 (1.3 g, 4.55 mmol) was dissolved in 13 mL of 80% aqueous acetic acid and the resulting solution was warmed to 60°C and stirred for 5 h. The solution was then concentrated under vacuum to furnish an oily crude residue that was purified by flash chromatography eluting with EtOAc/MeOH 85:15. There was obtained a pure terminal diol intermediate (804 mg, 72%) as a colorless oil: ¹H NMR (300 MHz, CDCl₃, 75:25 mixture of isomers) δ 5.42 (m, 0.75H), 5.32 (m, 0.25H), 4.54 (m, 0.25H), 4.50 (bd, J=10.5 Hz, 0.75H), 3.95 (m, 0.75H), 3.3-3.8 (m, 5.25H), 3.01(td, J=13.3, 3.2 Hz, 0.75H), 2.49 (td, J=13.3, 2.9 Hz, 0.25H), 2.12 (s, 3×0.75H), 2.07 (s, 3×0.25H),2.01 (s, 3×0.25 H), 1.98 (s, 3×0.75 H), 1.5–1.9 (m, 4H). The diol was then dissolved in CH₂Cl₂ (25 mL) and treated with a 0.65 M aqueous NaIO₄ solution (25 mL) and chromatography grade SiO₂ (20 g). The resulting heterogeneous mixture was vigorously stirred at room temperature for 20 min, after which time the slurry was filtered under suction and the silica thoroughly washed with CH₂Cl₂. The filtrates were evaporated to afford aldehyde 14 (664 mg, 95%) as a colorless oil: R_f =0.5 (EtOAc/MeOH 90:10); $[\alpha]_D^{20} = -40.9$ (c=0.7, CHCl₃); ¹H NMR (300 MHz, CDCl₃, 80:20 mixture of isomers, major isomer) δ 9.51 (s, 1H), 5.48 (m, 1H), 5.42 (m, 1H), 3.80 (bd, J=13.4 Hz, 1H), 3.15 (td, J=12.0, 3.1 Hz, 1H), 2.19 (s, 3H), 2.02 (s, 3H), 1.5–2.0 (m, 4H); ¹³C NMR (75 MHz, CDCl₃, major isomer) δ 196.9, 170.8, 169.8, 65.0, 61.9, 44.3, 25.8, 20.9, 20.7, 19.8.

(2R,3R)-1,3-N,O-Diacetyl-2-formyl-3-hydroxypiperidine ent-14

The title compound was prepared following the two-step procedure described to transform 13 into 14, starting with 1.2 g (4.2 mmol) of piperidine *ent*-13. There were obtained 628 mg (70%) of aldehyde *ent*-14 as an oil: R_f =0.5 (EtOAc/MeOH 90:10); [α]_D²⁰=+39.0 (c=0.5, CHCl₃). The ¹H and ¹³C NMR spectra coincided with those of its enantiomer 14.

(2S,3S)-N,O-1,3-Diacetyl-3-hydroxypipecolic acid 15

Aldehyde 14 (650 mg, 3.04 mmol) was dissolved in 25 mL of a CCl₄–MeCN–acetone–H₂O solvent mixture (1:1:0.3:1.4 ratio) and treated with 1.0 g of solid NaIO₄ and a catalytic amount of RuO₂·2H₂O (20 mg). The resulting mixture was stirred at room temperature for 1 h, quenched with propan-2-ol and filtered on a Celite pad. The filtrate was evaporated and the residue was purified by flash chromatography eluting with EtOAc/MeOH/30% aq. NH₃ 6:4:0.3 to furnish 543 mg (78%) of protected pipecolic acid 15 as a foamy solid: R_f=0.4 (EtOAc/MeOH/30% aq. NH₃ 6:4:0.3); $[\alpha]_D^{20}$ =-36.7 (c=0.6, CHCl₃); ¹H NMR (300 MHz, CDCl₃, 70:30 mixture of isomers, major isomer) δ 7.67 (bs, 1H), 5.39 (m, 1H), 5.09 (m, 1H), 3.65 (bd, *J*=12.8 Hz, 1H), 3.22 (bt, *J*=12.0 Hz, 1H), 2.08 (s, 3H), 1.95 (s, 3H), 1.4–1.9 (m, 4H); ¹³C NMR (75 MHz, CDCl₃, major isomer) δ 173.0, 172.0, 170.8, 69.2, 56.9, 43.8, 25.3, 21.5, 21.2, 21.1. Anal. Calcd for C₁₀H₁₅NO₅: C, 52.40; H, 6.60; N, 6.11. Found: C, 52.51; H, 6.49; N, 6.03.

(2R,3R)-1,3-N,O-Diacetyl-3-hydroxypipecolic acid ent-15

The title compound was prepared starting with 600 mg (2.81 mmol) of aldehyde *ent-14* according to the procedure described for 15. There were obtained 510 mg (79%) of protected pipecolic acid *ent-15* as a foamy solid: R_f =0.4 (EtOAc/MeOH/30% aq. NH₃ 6:4:0.3); [α]_D²⁰=+35.9 (c=0.5, CHCl₃). The ¹H and ¹³C NMR spectra coincided with those of its enantiomer 15. Anal. Calcd for $C_{10}H_{15}NO_5$: C, 52.40; H, 6.60; N, 6.11. Found: C, 52.28; H, 6.70; N, 5.98.

(2S,3S)-3-Hydroxypipecolic acid L-1

To a solution of protected pipecolic acid 15 (520 mg, 2.27 mmol) in 30 mL of methanol were added 10 mL of a 1% methanolic NaOMe solution portionwise until complete disappearance of the starting material (about 3 h). The reaction mixture was then concentrated in vacuum to give a crude residue that was purified by flash chromatography eluting with EtOAc/MeOH/30% aq. NH₃ (6:4:0.3). There was obtained a partially *O*-deprotected pipecolic acid intermediate (382 mg, 90%) as a gummy solid: 1 H NMR (300 MHz, CD₃OD, 50:50 isomeric mixture) δ 5.05 (m, 0.5H), 4.50 (m, 1H), 4.40 (bd, J=12.7 Hz, 0.5H), 4.30 (m, 0.5H), 3.74 (bd, J=13.0 Hz, 0.5H), 3.40 (td, J=13.0, 3.9 Hz, 0.5H), 2.85 (td, J=13.0, 3.0 Hz, 0.5H), 2.14 (s, 3×0.5H), 2.05 (s, 3×0.5H), 1.6–1.8 (m, 2H), 1.3–1.5 (m, 2H).

This intermediate (382 mg, 2.04 mmol) was dissolved in a 6 N aqueous HCl solution (10 mL) and the resulting mixture gradually warmed to 80°C. After being stirred at this temperature for 10 h, the solution was concentrated under vacuum to give a crude residue that was purified by flash chromatography eluting with EtOAc/MeOH/30% aq. NH₃ (5:5:0.7). Pure pipecolic acid L-1 was recovered (255 mg, 86%) as an off-white solid material: mp 230–238°C (decomp.); R_f =0.32 (EtOAc/MeOH/30% aq. NH₃ 5:5:1); $[\alpha]_D^{20}$ =+12.9 (c=0.23, 10% aq. HCl) {lit.^{4a} $[\alpha]_D^{20}$ =+13 (c=0.4, 10% aq. HCl); lit.^{4b} $[\alpha]_D^{20}$ =+15 (c=0.5, 10% aq. HCl)}; ¹H NMR (300 MHz, D₂O) δ 4.05 (ddd, J=10.5, 7.2, 3.0 Hz, 1H, H-3), 3.49 (d, J=7.2 Hz, 1H, H-2), 3.27(m, 1H, H-6_{eq}), 2.98 (ddd, J=12.9, 9.0, 3.3 Hz, 1H, H-6_{ax}), 1.91 (m, 2H, H-4_{eq}, H-5_{eq}), 1.64 (m, 2H, H-4_{ax}, H-5_{ax}); ¹³C NMR (75 MHz, CD₃OD) δ 176.4, 67.3, 62.9, 43.9, 30.2, 20.1. Anal. Calcd for $C_6H_{11}NO_3$:C, 49.65; H, 7.64; N, 9.65. Found: C, 49.77; H, 7.80; N, 9.52.

(2R,3R)-3-Hydroxypipecolic acid D-1

The title compound was prepared by starting with 500 mg (2.18 mmol) of protected pipecolic acid *ent-***15** by paralleling the two-step procedure described to transform **15** into L-1. After chromatographic purification (flash chromatography, EtOAc/MeOH/30% aq. NH₃ 5:5:0.7), there was obtained pipecolic acid D-1 (237 mg, 75%) as a white solid material: mp 234–239°C (decomp.); R_f =0.32 (EtOAc/MeOH/30% aq. NH₃ 5:5:1); $[\alpha]_D^{20}$ =-13.0 (c=0.2, 10% aq. HCl) {lit.^{4a} $[\alpha]_D^{20}$ =-14 (c=0.5, 10% aq. HCl)}. The ¹H NMR and ¹³C NMR spectral data were identical to those reported for its enantiomer L-1. Anal. Calcd for $C_6H_{11}NO_3$:C, 49.65; H, 7.64; N, 9.65. Found: C, 49.52; H, 7.80; N, 9.77.

Structure determination of compound 12

Crystal data: $C_{10}H_{18}NO_3$, M=200.2, triclinic, space group P1, a=9.769(5), b=5.535(2), c=5.196(2) Å, $\alpha=103.31(1)$, $\beta=77.68(1)$, $\gamma=97.28(1)^\circ$, V=266.2(2) Å³, Z=1, F(000)=109, Dc=1.249 Mg m⁻³, $\mu(Cu-K\alpha)=7.5$ cm⁻¹. The intensity data were collected at room temperature on a Siemens AED diffractometer over a 3–70° θ range; h, –11 to 11; k, –6 to 6; l, 0 to 6, by using the Cu-K α radiation ($\lambda=1.54178$ Å) and $\theta-2\theta$ scanning. Of the 1012 unique data measured, 589 had $I>2\sigma(I)$ and were used in the subsequent structural solution. The data were corrected for Lorentz and polarization effects, but not for absorption in view of small crystal dimensions ($0.43\times0.47\times0.70$ mm). The structural determination was carried out by SIR92. ^{13a} The structure was then refined by full-matrix least-squares methods (SHELXL-96)^{13b} using anisotropic temperature factors for all the non-hydrogen atoms. All H-atoms were calculated at idealized positions with the exception of the hydrogen of the piperidine nitrogen which was located with a difference Fourier map and not refined. At convergence, the discrepancy indices R and wR were 0.066 and 0.154, respectively ($w=1/[\sigma^2(F_o^2)+(0.1360P)^2+0.0P]$ where $P=(Max(F_o^2,0)+2F_o^2)/3$. Scattering factors for C, H, N and O were taken from the literature. ^{13c} Molecular-geometry calculations were carried out by using the computer program PARST^{13d} and the structure drawing by using the ORTEP and PLUTON program. ^{13e,f} Lists of refined coordinates,

e.s.d.'s, and thermal parameters obtained from the crystallographic analysis have been deposited at the Cambridge Crystallographic Data Centre (CCDC).

Acknowledgements

We are grateful to Ministero dell'Università e della Ricerca Scientifica e Tecnologica (MURST), Italy, and to Consiglio Nazionale delle Ricerche (CNR), Italy, for the financial support and to Centro Interdipartimentale Misure "G. Casnati" for NMR facilities.

References

- Romeo, J. T.; Swain, L. A.; Bleecker, A. B. Phytochemistry 1983, 22, 2687. Schenk, V. W.; Schutte, H. R. Flora 1963, 153, 426. Vanderhaeghe, H.; Janssen, G.; Compernolle, F. Tetrahedron Lett. 1971, 28, 2687. Gillard, G.; Abraham, A.; Anderson, P. C.; Beaulieu, P. L.; Bogri, T.; Bousquet, Y.; Grenier, L.; Guse, I.; Lavallée, P. J. Org. Chem. 1996, 61, 2226.
- 2. Bashyal, B. P.; Chow, H.-F.; Fellows, L. E.; Fleet, G. W. J. Tetrahedron 1987, 43, 415.
- 3. Niwa, T.; Tsuruoka, T.; Inouye, S.; Naito, Y.; Koeda, T.; Niida, T. J. Biochem. 1972, 72, 207.
- Previous syntheses of trans- and cis-3-hydroxypipecolic acids: (a) Greck, C.; Ferreira, F.; Genêt, J. P. Tetrahedron Lett. 1996, 37, 2031. (b) Agami, C.; Couty, F.; Mathieu, H. Tetrahedron Lett. 1996, 37, 4001. (c) Roemmele, R. C.; Rapoport, H. J. Org. Chem. 1989, 54, 1866. (d) Knight, D. W.; Lewis, N.; Share, A. C.; Haigh, D. Tetrahedron: Asymmetry 1993, 4, 625.
- Schöllkopf, U.; Neubauer, H.-J.; Hauptreif, M. Angew. Chem. Int. Ed. Engl. 1985, 24, 1066.
 Williams, R. M.; Sinclair, P. J.; Zhai, D.; Chen, D. J. Am. Chem. Soc. 1988, 110, 1547. Agami, C.;
 Couty, F.; Prince, B.; Puchot, C. Tetrahedron 1991, 47, 4343. Schickli, C. P.; Seebach, D. Liebigs Ann. Chem. 1991, 655. Enders, D.; Funk, R.; Klatt, M.; Raabe, G.; Hovesteyde, E. R. Angew. Chem. Int. Ed. Engl. 1993, 32, 418.
- Selected works exploiting 2-silyloxy furan and related reagents: (a) Spanu, P.; Rassu, G.; Ulgheri, F.; Zanardi, F.; Battistini, L.; Casiraghi, G. Tetrahedron 1996, 52, 4829. (b) Zanardi, F.; Battistini, L.; Nespi, M.; Rassu, G.; Spanu, P.; Cornia, M.; Casiraghi, G. Tetrahedron: Asymmetry 1996, 7, 1167. (c) Soro, P.; Rassu, G.; Spanu, P.; Pinna, L.; Zanardi, F.; Casiraghi, G. J. Org. Chem. 1996, 61, 5172. (d) Rassu, G.; Zanardi, F.; Battistini, L.; Gaetani, E.; Casiraghi, G. J. Med. Chem. 1997, 40, 168. (e) Rassu, G.; Pinna, L.; Spanu, P.; Zanardi, F.; Battistini, L.; Casiraghi, G. J. Org. Chem. 1997, 62, 4513.
- 7. The actual experimental conditions in the coupling reaction resulted after an extensive optimization procedure. Of note, when stoichiometric BF₃·OEt₂ (CH₂Cl₂, -80°C) or catalytic TMSOTf (CH₂Cl₂, -80°C), InCl₃ (THF, 0°C), Yb(OTf)₃ (CH₂Cl₂, 0°C) were used, a decrease in efficiency and/or diastereoselectivity was observed, being compounds 8 and 9 flanked by two other isomers (diastereomeric ratios ranging from 60:40 to 80:20).
- 8. When 8 was treated with Et₃N in CH₂Cl₂, no base-promoted interconversion into 9 was observed, thus confirming that these compounds were not epimeric at C-4. The 4,5-anti-stereodisposition within 9 was deduced by the ¹H NMR spectrum of an advanced trans-configured piperidine intermediate related to compound 12.
- 9. As a rule, lactones having 4R absolute configuration are dextrorotatory, while those having 4S absolute configuration are levorotatory. With few exceptions, this empirical guideline proved widely applicable also to a variety of closely related substances. Casiraghi, G.; Colombo, L.; Rassu, G.; Spanu, P.; Gasparri Fava, G.; Ferrari Belicchi, M. Tetrahedron 1990, 46, 5807.
- 10. Burgess, L. E.; Meyers, A. I. J. Org. Chem. 1992, 57, 1656.
- 11. Mukaiyama, T.; Narasaka, K.; Banno, K. Chem. Lett. 1973, 1011. Mukaiyama, T.; Narasaka, K.; Banno, K. J. Am. Chem. Soc. 1974, 96, 7503.

- (a) Berkovitch-Yellin, Z.; Leiserowitz, L. Acta Crystallogr. 1984, B40, 159. (b) Taylor, R.; Kennard, O. J. Am. Chem. Soc. 1982, 104, 5063. (c) Cremer, D.; Pople, J. A. J. Am. Chem. Soc. 1975, 97, 1354. (d) Kodoma, Y.; Tsuruoka, T.; Niwa, T.; Inouye, S. J. Antibiot. 1985, 38, 116. (e) Evrard, G.; Durant, F.; Marlier, M. Cryst. Struct. Comm. 1974, 3, 61. (f) Nardelli, M. Acta Crystallogr. 1983, C39, 1259. (g) Casiraghi, G.; Ulgheri, F.; Spanu, P.; Rassu, G.; Pinna, L.; Gasparri Fava, G.; Belicchi Ferrari, M.; Pelosi, G. J. Chem. Soc., Perkin Trans. 1 1993, 2991 and references therein.
- (a) Altomare, A.; Burla, M. C.; Camalli, M.; Cascarano, G.; Giacovazzo, C.; Guagliardi, A.; Polidori, G. J. Appl. Crystallography 1994, 27, 435. (b) Sheldrick, G. M. SHELXL-96, a program for crystal-structure refinement, University of Goettingen, 1996. (c) International Tables for X-Ray Crystallography; Kynoch Press: Birmingham, 1975; p. 4. (d) Nardelli, M. Comput. Chem. 1983, 7, 95. (e) Johnson, C. K. ORTEP Report ORNL 3794; Oak Ridge National Laboratory: Tennesee, 1965. (f) Spek, A. L. Acta Crystallogr. 1990, A46, 194.

(Received in UK 21 July 1997)